Determining water extractable P in animal manure and biosolids Ann Wolf, Pennsylvania State University Philip Moore, Jr., USDA-ARS, Fayetteville, AR Peter Kleinman, USDA-ARS, University Park, PA Dan Sullivan, University of Oregon

Introduction:

The water extractable P content of land-applied manures and biosolids is a key water quality indicator, and has been used in a variety of contexts to evaluate different materials on the basis of their potential to release dissolved P to runoff water. Consistent and meaningful measurement of water extractable P has been obtained by a variety of methods, each with its own set of advantages and limitations(Self-Davis and Moore, 2000; Sharpley and Moyer, 2000, Haggard et al., 2005; Baum et al., 2006; Vadas and Kleinman, 2006). Key differences in methods include how materials are processed (drying, grinding), whether fresh materials are extracted on a dry weight equivalent basis or on a wet basis, the amount of sample that is extracted, extraction ratio (solution:solids), extraction time, solid separation technique, and method of P determination (ICP vs. colorimetry). Recent efforts by SERA-17 culminated in the selection of a universal water extraction protocol that addresses the concerns of testing laboratories (reproducibility, ease of implementation, and adaptation to different manures and biosolids) while providing a strong correlation between WEP and with dissolved P in runoff (Kleinman et al., 2007).

Two water extraction methods are reported in this chapter. The first, adapted from Kleinman et al. (2007) represents the consensus method of SERA-17 and, more importantly, has been shown to work well with a broad array of manures and biosolids (dry and liquid). The second method, derived from Self-Davis and Moore (2000), is also reported because it is currently required by the Arkansas P Index for Pastures. That method has been used consistently and reliably with poultry litter samples. However, because the Self-Davis and Moore (2000) method does not fix the extraction ratio, it does not provide reliable comparisons of water extractable P across manures with varying moisture content (see Kleinman et al., 2002). Furthermore, due to the relatively low extraction ratio (approximately 10:1) of the Self-Davis and Moore (2000) method, it presents practical difficulties and cannot be used with certain manures and biosolids (see Kleinman et al., 2007).

Universal Water Extractable P Test for Manure and Biosolids (adapted from Kleinman et al., 2007)

Summary of Method:

A representative sample containing 2.0 g of solids is extracted with water at a 100:1 solution:solids ratio for 1 hr, centrifuged and, if necessary, filtered. The filtrate is analyzed for phosphorus (P) by inductively coupled argon plasma spectroscopy (ICP).

Apparatus and Materials:

1. Analytical balance, 300 g capacity, minimum accuracy of \pm 0.001 g

- 2. 250 mL centrifuge bottles
- 3. Reciprocating shaker capable of 180-200 epm
- 4. Centrifuge capable of 3,000 or greater rpm
- 5. Whatman No. 40 filter paper and filter funnels

Reagents:

- 1. Reagent grade water, minimum resistance of $17 \text{ M}\Omega \text{ cm}$
- 2. ICP Standards: P standards ranging from 0 (Reagent grade water blank) up to 100 mg/L P.

Sample Preservation and Handling :

- 1. All sample containers should be pre-washed with phosphate-free detergents and rinsed with deionized water. Plastic and glass containers are both suitable.
- 2. Samples shall be refrigerated (4^o C) upon receipt and analyzed within three weeks.

Procedure

- 1. Determine the percent solids of the manure on a separate subsample (Peters, 2003).
- 2. Based on the percent solids determination, weigh or pipet a sample containing 2.0 g solids (see calculations below) into a 250 mL centrifuge bottle.
- 3. Add reagent grade water to bring to a final weight of 202 g. If solids content of sample is less than or equal to 1%, do not add additional water. Use 200 g of the asreceived sample.
- 4. Shake samples on a reciprocating shaker (180-200 epm) for 60 minutes.
- 5. Remove bottles from shaker and centrifuge (3,000 rpm or greater) for 10 minutes. If particulates are present, filter the centrifugate (No. 40 Whatman).
- 6. Analyze centrifugate for P on the ICP. If sample extract can not be analyzed immediately, acidify to prevent precipitation of calcium phosphates by adding 5 drops of concentrated HCl for each 20 mL of extract to lower pH to approximately 2.0. Acidified extracts can be held for up to three weeks before P measurement. *Samples may also be analyzed colorimetrically but should be referenced to ICP analyses due to matrix interference issues.*

Calculations:

1. Sample size needed to provide 2.0 g solids is determined as follow:

Sample size to provide 2.0 g = 2 x
$$\frac{100}{\%}$$
 solids

2. The P concentration on a dry weight basis in % is determined as follows:

P (%) dry weight basis =
$$\frac{\text{ICP P result (mg/L) x 0.02}}{\text{Sample size wet (g)}} \times \frac{100}{\% \text{solids}}$$

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